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Grape seed and skin extract protects against acute chemotherapy toxicity induced by doxorubicin in rat red blood cells and plasma

Grape seed and skin extract protects against acute chemotherapy toxicity induced by doxorubicin in rat red blood cells and plasma

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Abstract

In this study, the protective role of grape seed and skin extract (GSSE) against doxorubicin-induced blood toxicity has been evaluated in rats. Rats were treated with the extract for 8 days and injected with doxorubicin (20 mg/kg) at the 4th day. At the end of the treatment, blood samples were collected for oxidative stress parameters determination and anti-oxidant enzymes. Doxorubicin increased erythrocytes and plasma malondialdehyde, free iron, H₂O₂ and carbonylation, decreased calcium and also decreased erythrocytes catalase, peroxidase and superoxide dismutase (specially the Fe isoform). Doxorubicin also decreased plasma catalase and superoxide dismutase (Cu/Zn and Fe isoforms) but increased peroxidase. Doxorubicin increased plasma alanine aminotransferase and aspartate aminotransferase but decreased them within erythrocytes. GSSE co-treatment counteracted almost all deleterious effects induced by doxorubicin. In conclusion, doxorubicin induced a pro-oxidative stress into rat erythrocytes and plasma and GSSE exerted anti-oxidant properties which can be attributed to free iron and calcium modulation.

Introduction

Doxorubicin (Dox) (Adriamycin) has been used in oncologic practice since the late 1960s. It held promise as a powerful drug in the fight against cancer (Singal et al., 1998). However, the clinical efficacy of this drug is limited due to damages toxicity in adults and pediatric cancer patients (Buzdar et al., 1985). The exact causal mechanism of Dox induced toxicity remains unclear, but most of the evidence indicates that free radicals are involved (Singal et al., 1987; Sinha et al., 1987a). The chemical structure of Dox is prone to the generation of free radicals, (Sinha et al., 1987b) and the oxidative stress that results correlates with cellular injury (Rosen and Halpern, 1990). In addition, Dox administration is

associated with a decrease in endogenous anti-oxidants responsible for the scavenging of free radicals (Singal et al., 1995) leading to increased oxidative stress, which is followed by damages in organism (Doroshov et al., 1980).

Grape seed and skin extract (GSSE) is a nutritional supplement exhibiting beneficial health effects (Suwanaphet et al., 2010). GSSE is a complex mixture of polyphenolics classified as flavonoid and non flavonoid compounds (Khanal et al., 2009). Flavonoids, which are highly concentrated in grape seeds are mainly composed of monomeric catechins, proanthocyanidins and flavonols, such as quercetin (Renaud and De Lorgeril, 1992). Non flavonoids, highly abundant in grape skin,



contain stilbenes such as resveratrol, which is at the basis of the french paradox (Renaud and De Lorgeril, 1992). GSSE has wide-ranging benefits including cardioprotective (Decordé et al., 2009), renoprotective (Safa et al., 2010), and neuroprotective (Wang et al., 2009) effects. GSSE also protects against hepatic ischemia-reperfusion injury (Sehirli et al., 2008), biliary obstruction (Dulundu et al., 2007), and azathioprine-induced hepatotoxicity in rats (El-Ashmawy et al., 2010). More specifically, resveratrol is multi-organ protective owing to its anti-oxidant (Kelen and Tepe, 2007) and anti-inflammatory properties (Kowalczyk et al., 2010). Proanthocyanidins exert antineoplastic effects by inducing cytotoxicity towards some cancer cells (Chatelain et al., 2011), by cell cycle arrest and induction of apoptosis (Kaur et al., 2008). In addition quercetin and rutin are neuroprotective (Jimenez-Aliaga et al., 2011), and catechin, epicatechin, and gallic acid have exhibited cholesterol-lowering activity (Ngamukote et al., 2011).

In the present work, we evaluated the effect of an acute administration of Dox on oxidative stress induced in the blood compartment as well as the putative protection offered by GSSE. Dox induces a pro-oxidative state and GSSE exerts potent anti-oxidative properties by counteracting the Fenton reaction of H₂O₂ with free iron, leading to intracellular calcium dysregulation.

Materials and Methods

Reagents

2-thiobarbituric acid (TBA); 2,6,-di-tert-butyl-4-hydroxy-toluene (BHT); trichloroacetic acid (TCA); hydrogen peroxide (H₂O₂); 2-methoxyphenol (gaiacol); bovine catalase and 4-(1-Hydroxy-2-methylamino-ethyl)-benzene-1,2-diol (epinephrine), 2,4-dinitrophenyl hydrazine (DNPH) were obtained from Sigma-Aldrich (Germany). Doxorubicin hydrochloride was purchased from Pharmacia Italia (Italy).

Preparation of grape seed and skin extract

GSSE was processed from a grape cultivar (Carignan) of *Vitis vinifera* from northern Tunisia. Waste from winemaking was collected from Tardi Cooperative Winery (Ain Ghelal). Seeds and skin were dried and ground separately with an electric mincer (FP3121 Moulinex) until a fine powder was obtained. Total phenolic content was determined by the folin-Ciocalteu colorimetric method (Singleton and Rossi, 1965), flavonoids and condensed tannins according to Dewanto et al. (2002) and Sun et al. (1998) respectively (Table I). Powder mixture containing grape seed (50%) and skin (50%) was dissolved in 1 mL of 10% ethanol in the dark, vigorously vortexed for 10 min, centrifuged at 10,000 g for 15 min at 4°C for debris elimination and the

supernatant containing soluble polyphenols was used.

Animals and treatment

Female Wistar rats (220-40 g) were used in these experiments in accordance with the ethic committee of Tunis University for care and use of animals in conformity with NIH guideline (NIH, 1985). They were provided with food and water ad libitum and maintained in animal facility at fixed temperature of 22 ± 2°C with a 12 hours light-dark cycle. Rats were randomly divided into four groups of six animals each and daily intraperitoneally injected with either vehicle (C: 10% ethanol) or GSSE (500 mg/kg *bw*) for 8 days. Dox was dissolved in saline and administered at 20 mg/kg *bw* by a single intraperitoneal injection on the fourth day. At the end of the experiment, rats were anesthetized with 0.5 mL urethane (40 mg/mL) and sacrificed, their blood collected and processed for plasma and erythrocytes parameters using automates and biochemical determination of anti-oxidant status parameters.

Blood processing

Whole blood was obtained by cardiac puncture and collected into heparinized tubes. Erythrocytes were isolated from plasma by centrifugation at 1000 g for 10 min at 4°C and homogenized using a hypotonic buffer Tris-HCl 10 mM pH 7.5, MgCl₂ 5 mM, NaCl 10 mM.

Plasma and erythrocytes transaminases measurement

Plasma and erythrocytes aspartate aminotransferase (AST) and alanine aminotransferase (ALT) were measured enzymatically using a commercial kit from Biomaghreb (Ariana, Tunisia) following the manufacturer's procedure (Reitmans and Frankels, 1957).

Lipoperoxidation

Lipoperoxidation was determined by malondialdehyde (MDA) measurement according to the double heating method (Draper and Hadley, 1990). Briefly, aliquots from erythrocyte homogenates or plasma were mixed with BHT-TCA solution containing 1% BHT (m/v) dissolved in 20% TCA (m/v) and centrifuged at 1000 g for 5 min at 4°C. The supernatant was blended with 0.5N HCl, and 120 mM TBA in 26 mM Tris, and then heated at 80°C for 10 min. After cooling, absorbance of the resulting chromophore was determined at 532 nm using a Bio-Rad UV-visible spectrophotometer. MDA levels were determined by using an extinction coefficient for MDA-TBA complex of 1.56 10⁵ M⁻¹cm⁻¹.

Protein carbonylation

Oxidative damage to proteins was evaluated by quantifying protein carbonylation in red blood cells homogenates or plasma according to Levine et al. (1990). After protein were precipitated with 20% TCA and centrifuged at x11,000 g for 3 min at 4°C (Beckman J20),

the pellet was dissolved in 10 mM DNPH-containing buffer. After 3 washes with ethanol- ethylacetate (1:1), the pellet was dissolved in 20 mM potassium phosphate (pH 2.3) containing 6M guanidine chloride and absorbance measured at 366 nm using the molar extinction coefficient of 22000 M-1cm-1. Results were expressed as nmol carbonyl residues/mg protein.

Protein determination

Total soluble proteins were determined according to the biuret method (Ohnishi and Barr, 1978). Briefly, at acidic pH, soluble proteins constituted with copper a colourful complex measurable at 546 nm using a SmartSpec 3000 Bio-Rad UV-visible spectrophotometer (Germany).

Anti-oxidant enzyme activity assays

All spectrophotometric analyses of erythrocytes and plasma anti-oxidant enzyme activities were performed with a SmartSpec 3000 Bio-Rad UV-visible spectrophotometer (Germany).

Catalase (CAT) activity was assayed by measuring the initial rate of H₂O₂ disappearance at 240 nm (Aebi, 1984). The reaction mixture contained 33 mM H₂O₂ in 50 mM phosphate buffer (pH 7.0) and erythrocytes extract or plasma. CAT activity was calculated using an extinction coefficient of 40 mM-1cm-1 for H₂O₂.

Peroxidase (POD) activity was measured at 25°C using guaiacol as the hydrogen donor. The reaction mixture contained 9 mM guaiacol, 19 mM H₂O₂ in 50 mM phosphate buffer pH 7 and erythrocytes extract or plasma in a final volume of 1 mL. The reaction was initiated by the addition of H₂O₂ and monitored by measuring the increase in absorbance at 470 nm every 30 sec for 3 min. POD activity was expressed as nmol of guaiacol oxidized per min and calculated using a molecular extinction coefficient of 26.2 mM-1 (Chance and Maehly, 1955).

Superoxide dismutase (SOD) activity was determined by using a modified epinephrine assay (Misra and Fridovich, 1972). At alkaline pH, superoxide anion (O₂⁻) causes the auto-oxidation of epinephrine to adrenochrome. One unit of SOD is defined as the amount of extract that inhibits the rate of adrenochrome formation by 50%. Enzyme extract was added to a 2 mL reaction mixture containing 10 µL bovine catalase (0.4 U/µL), 20 µL epinephrine (5 mg/mL) and 62.5 mM sodium carbonate-sodium bicarbonate buffer (pH 10.2). Changes in absorbance were recorded at 480 nm. Characterization of SOD isoforms was performed using KCN (3 mM), which inhibits Cu/Zn-SOD or H₂O₂ (5 mM), affecting both Cu/Zn-SOD and Fe-SOD. Mn-SOD was insensitive to both inhibitors.

Free iron determination

Erythrocytes and plasma free iron levels were deter-

mined according to Leardi et al. (1998) using a commercially available kit from Biomaghreb. Briefly, at acidic pH 4.8 all Fe³⁺ released from transferrin were reduced by ascorbic acid into Fe²⁺, which constitutes with ferrozine a purple colourful complex measurable at 560 nm. Briefly, 50 µL of erythrocytes extract or plasma was added to 250 µL of reaction mixture containing ascorbic acid (5 g/L) and ferrozine (40 mM), and incubation was performed at 37°C for 10 min.

H₂O₂ determination

Erythrocytes and plasma H₂O₂ levels were determined enzymatically according to Kakinuma et al. (1979) using a commercially available kit from Biomaghreb. Briefly, in the presence of POD, H₂O₂ reacts with 4-amino-antipyrene and phenol to give a red-colored quinonimine that absorbs at 505 nm. Results are expressed as mmol H₂O₂/mg protein.

Calcium measurement

Intracellular calcium was determined according to Stern and Lewis (1957) using a commercially available kit from Biomaghreb. At basic pH, calcium constitutes with cresolphthalein a purple colorful complex measurable at 570 nm. Briefly, 50 µL of erythrocytes extract or plasma was added to 650 µL of reaction mixture containing 2-amino-2-methyl 1-propanol buffer (500 mmol/L), cresolphthalein (0.62 mmol/L), and hydroxy-8 quino- leine (69 mmol/L). Incubation was carried out at room temperature for 5 min assuming the complex was stable for 1 hour.

Statistical analysis

Data were analyzed by unpaired Student's t-tests or 1-way analysis of variance (ANOVA) and expressed as means ± standard error of the mean (SEM). All statistical tests were 2-tailed, and results with a p value <0.05 were considered significant.

Results

Figure 1 shows the effect of Dox and GSSE either alone or in combination on erythrocytes (Figure 1A) and plasma (Figure 1B) lipoperoxidation and on erythrocytes (Figure 1C) and plasma (Figure 1D) protein carbonylation. Dox increased erythrocytes and plasma lipoperoxidation and carbonylation, and GSSE, which had no significant effect per se, counteracted all Dox-induced deleterious effects. Noticeably that plasma MDA increased significantly over control when animals were co-treated with Dox and GSSE (Figure 1B).

Dox significantly decreased erythrocytes ALT (Figure 2A) and AST (Figure 2C) and increased plasma ALT (Figure 2B) and AST (Figure 2D). Treatment with GSSE counteracted all Dox-induced disturbances to near control level except erythrocytes AST which increased

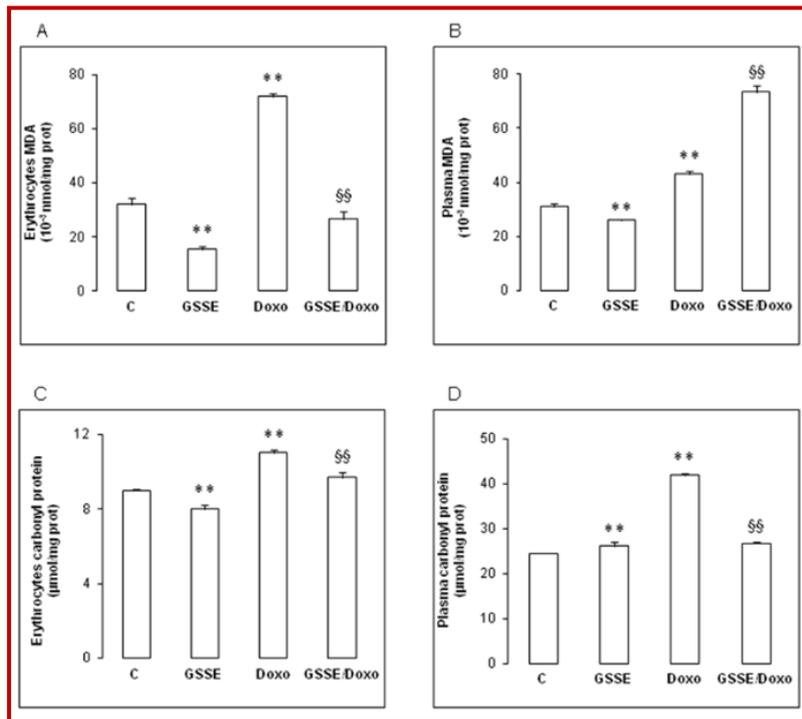


Figure 1: Effect of doxorubicin and GSSE on erythrocytes and plasma oxidation. Rats were pre-treated or not with GSSE during 8 days and challenged with a single dose of Dox at the fourth day. Erythrocytes (A) and plasma (B) MDA and erythrocytes (C) and plasma (D) protein carbonylation were determined. Results are expressed as mean \pm SEM (n = 6). **p<0.01 vs C. §§ p<0.01 vs Dox

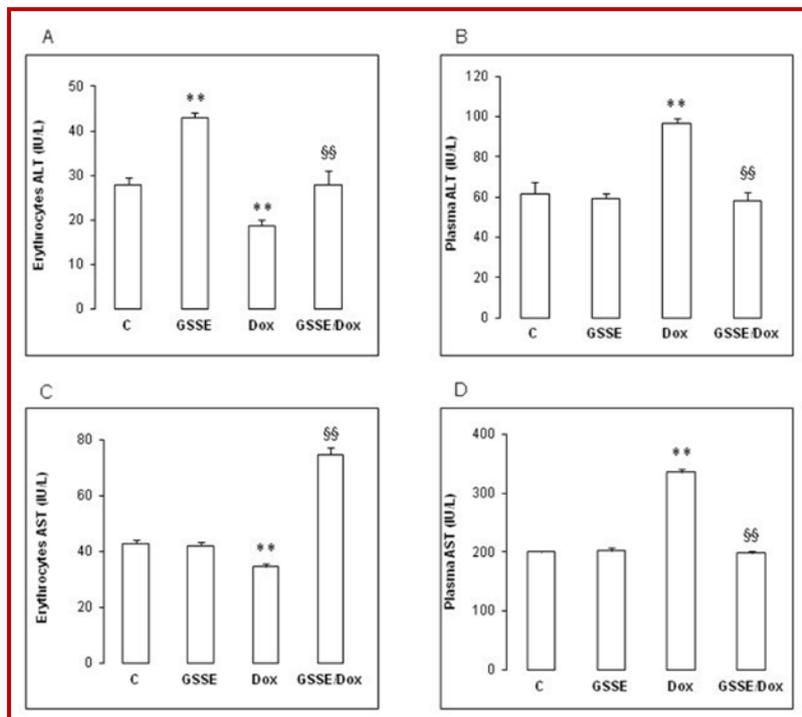


Figure 2: Effect of doxorubicin and GSSE on erythrocytes and plasma transaminases. Rats were pre-treated or not with GSSE for 8 days and challenged with a single dose of Dox at the fourth day. Erythrocytes (A) and plasma (B) ALT and erythrocytes (C) and plasma (D) AST were determined. Results are expressed as mean \pm SEM (n = 6). **p<0.01 vs C. §§ p<0.01 vs Dox

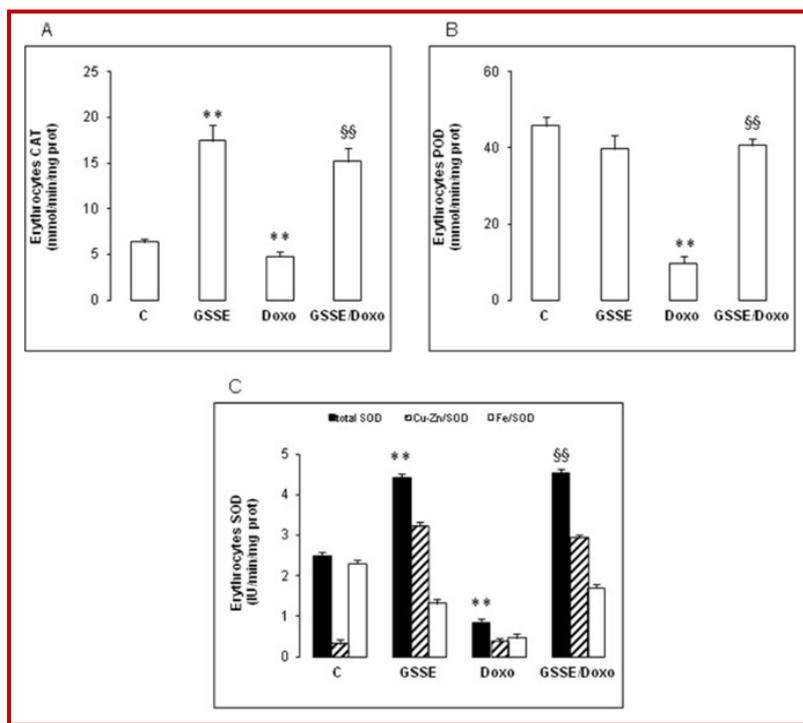


Figure 3: Effect of doxorubicin and GSSE on erythrocytes anti-oxidant enzyme activities. Rats were pre-treated or not with GSSE for 8 days and challenged with a single dose of Dox at the fourth day. Erythrocytes CAT (A), POD (B) and SOD (C) activities were determined. Results are expressed as mean \pm SEM (n = 6). **p<0.01 vs C. \$\$ p<0.01 vs Dox

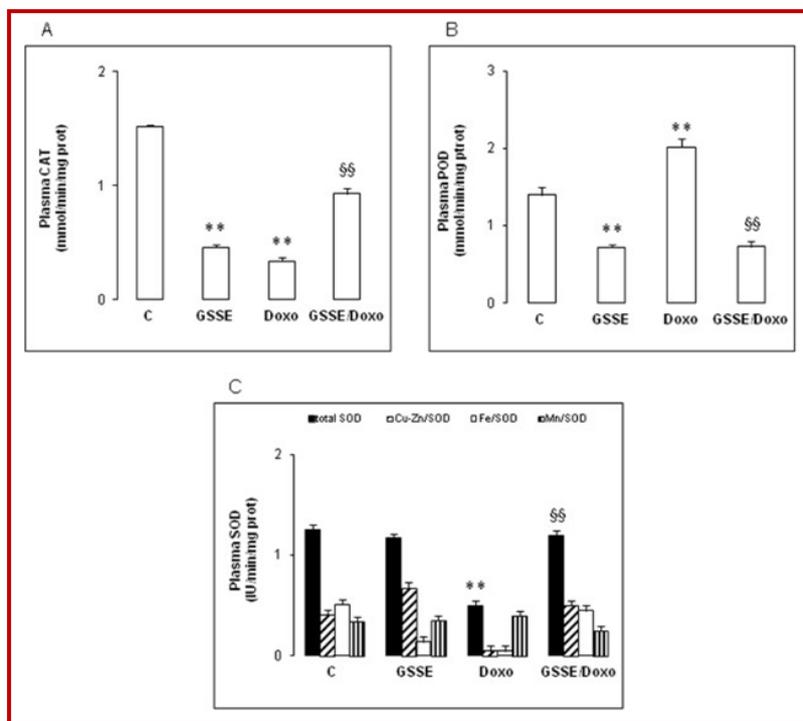


Figure 4: Effect of doxorubicin and GSSE on plasma anti-oxidant enzyme activities. Rats were pre-treated or not with GSSE for 8 days and challenged with a single dose of Dox at the fourth day. Plasma CAT (A), POD (B) and SOD (C) activities were determined. Results are expressed as mean \pm SEM (n = 6). **p<0.01 vs C. \$\$ p<0.01 vs Dox

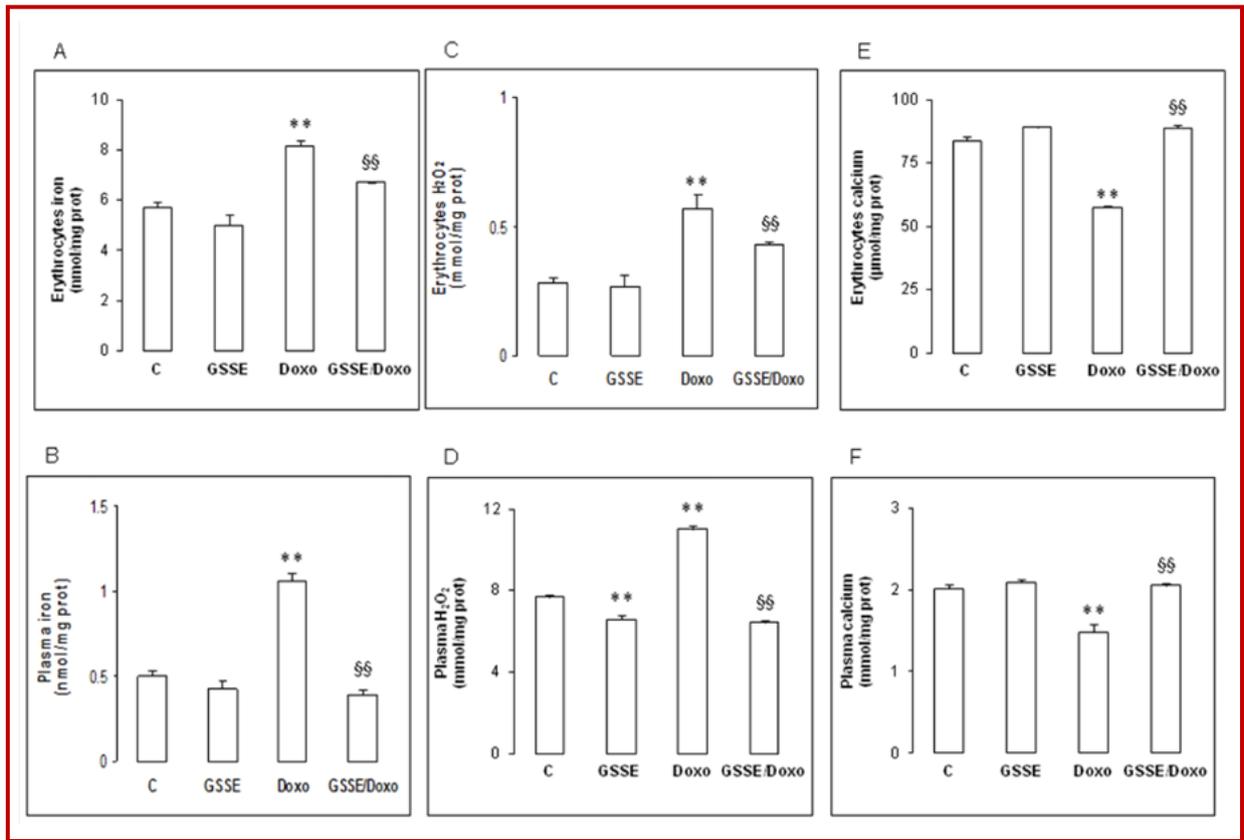


Figure 5: Effect of doxorubicin and GSSE on erythrocytes and plasma mediators

Rats were pre-treated or not with GSSE for 8 days and challenged with a single dose of Dox at the fourth day. Erythrocytes and plasma free iron (A, B), H_2O_2 (C, D) and calcium (E, F) were determined. Results are expressed as mean \pm SEM ($n = 6$). ** $p < 0.01$ vs C. §§ $p < 0.01$ vs Dox.

significantly following Dox and GSSE co-treatment.

Dox treatment down-regulated CAT (Figure 3A), POD (Figure 3B) and SOD (Figure 3C) activity (Figure 3A). Remarkably that among SOD Dox mainly affected the Fe isoform. GSSE alone had no effect on POD, increased CAT and SOD activities and in this latter case, mainly affected the Cu/Zn isoform. Importantly GSSE treatment counteracted all the deleterious effects of Dox on anti-oxidant enzyme activities to levels even higher than control i.e. CAT and Cu/Zn-SOD isoform.

The effect of Dox on plasma anti-oxidant enzyme activities is reported in Figure 4. Dox negatively affected CAT and SOD activities and in this latter case mainly depressed the Cu/Zn and Fe isoforms without affecting the Mn one. Pre-treatment with GSSE abrogated the Dox-induced disturbances in anti-oxidant enzyme activities to near control level.

We further sought to determine the putative involvement of intracellular mediators as Ca^{2+} in Dox-induced oxidative stress. In the same time, Dox increased free iron and H_2O_2 into erythrocytes and plasma and decreased free calcium. Once again, pre-treatment with

GSSE counteracted the Dox-induced blood disturbances of intracellular mediators.

Discussion

In the present study we analyzed the toxicity of Dox into the blood compartment. We found that Dox provoked a drastic oxidative stress status within erythrocytes and plasma as assessed by high MDA and carbonyl protein, elevated AST and ALT within plasma and a drastic depression of anti-oxidant enzymes in both compartments. Dox also increased free iron and H_2O_2 , it likely induced the highly toxic hydroxyl radical which in turn affects Ca^{2+} homeostasis (Ishii et al., 2006) and ultimately lead to erythrocyte death and anaemia. Several previous studies have shown the implication of Ca^{2+} disturbances in Dox-induced cardiotoxicity (Lebrecht et al., 2009). However to our knowledge our data are the first to describe such a phenomenon into the blood compartment.

Furthermore Dox treatment depressed CAT, POD and SOD activities within erythrocytes. Among SOD activi-

ties, the Fe-SOD was the only affected isoform, although we noticed in the same time that Dox increased both free iron and H₂O₂. Consequently Dox induced a drastic oxidative stress within erythrocytes that lead to hydroxyl radical generation which has been implicated in H₂O₂-induced calcium activation and cell death (Ishii et al., 2006). However in our present case we found Dox to depress erythrocyte Ca²⁺ (Jensen, 1986), whereas most studies described Dox-induced Ca²⁺ overloading (Kalivendi et al., 2005). Differential experimental conditions as Dox administration after a single high dose of 20 mg/kg could explain this discrepancy. Further studies should investigate which kind of calcium channels (L or T type) is involved in the mechanism of action of Dox using calcium channel modulators as verapamil or nifedipine.

Overall Dox-induced oxidative stress within erythrocytes appeared much more drastic than into the heart as demonstrated by some of us (Mokni et al., 2012). Because of the high polyunsaturated fatty acid content of their membranes and the high cellular concentration of oxygen, erythrocytes are highly susceptible to oxidative damage. Increased MDA level is known to affect the fluidity of the membrane lipid bilayer (Bryszewska et al., 1995) and high MDA level is generally correlated to stress-induced pathological conditions including aging (Rizvi and Maurya, 2007).

In our opinion, the most important result drawn from the present study is the ability of GSSE to prevent Dox-induced oxidative stress in rat erythrocytes and plasma compartment. GSSE is safe and devoid of any toxicity even at the high dosage used (Hebbar et al., 2005). GSSE protection could be mediated by ROS scavenging activity as demonstrated for resveratrol (Leonard et al., 2003).

GSSE extract is protective against cardiovascular diseases (Das and Maulik, 2006) and polyphenols containing GSSE as resveratrol and quercetin exerted anti-oxidant effects in human erythrocytes *in vitro* (Mikstacha et al., 2010). GSSE has been previously shown to protect multiple target organs from Dox (Mokni et al., 2012) and garlic-induced toxicity (Hamlaoui-Gasmi et al., 2012), owing to their anti-oxidant properties (Hamlaoui-Gasmi et al., 2011).

A possible mechanism by which GSSE could exert its beneficial effect on erythrocytes, is its ability to chelate free iron and to scavenge H₂O₂, re-establishing calcium homeostasis as recently found in rat heart subjected to a high fat regime (Charradi et al., 2011). Free iron is a well established catalyst of auto-oxidation and iron-mediated oxidations of cysteine residues represent a common mechanism through which H₂O₂ exerts its second messenger role in signal transduction pathways (Barbouti et al., 2007). Further studies should also analyze the implication of specific proteins involved in free iron metabolism as hepcidin (Isoda et al., 2010).

Moreover, H₂O₂ is able to induce dual roles in both survival and cell death pathways, largely depending on its concentration and also on the cell type. H₂O₂ exerted a prolongevity effect in several species including rat by induction of SOD activity (Yoshioka et al., 1994). GSSE could also exert its beneficial effects by its fatty acid moiety, having in mind the complex composition of GSSE (Khanal et al., 2009). Effective protection of blood cells could be achieved by some synergism between polyphenols and fatty acids containing GSSE. In this respect, fatty acids from magnolias seeds were recently shown to inhibit Dox-induced increase in intracellular Ca²⁺, ROS generation and apoptosis in rat cardiomyocytes (Park et al., 2008). Studies aiming at the identification of the bioactive components from GSSE responsible for the blood protection are in progress.

Another interesting feature of GSSE is its ability to increase erythrocyte CAT and SOD activities, more specifically the Cu/Zn isoform in this latter case. Increased activity in anti-oxidant enzymes could correspond to post-translational modifications as phosphorylation inducing some gain of function as reported by Borchi et al. (2010) on human heart failure about increased CAT and GPx activities after tyrosine phosphorylation. In our case, GSSE could induce phosphorylation of CAT and Cu/Zn-SOD and by this way enhanced their activities without modifying their expression nor abundance. Moreover we can't exclude a positive effect of GSSE on Cu or Zn accumulation into erythrocytes leading to increased activity of the corresponding SOD isoform. Our present data are consistent with those of Danz et al. (2009) who showed cardioprotective effect of resveratrol against Dox-induced oxidative damage partly by its effect on Mn-SOD activity but not on protein expression.

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Conflict of Interest

Authors declare no conflict of interest.

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