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Abstract

The main aim of the present research work was to demonstrate the antitumor and apoptotic activities of oxypeucedanin hydrate monoacetate isolated from *Angelica dahurica* against Caco-2 colon cancer cells. MTT cell viability assay along with clonogenic assays were used to study the effect of the compound on cell cytotoxicity and the colony forming tendency. Fluorescence microscopy using acridine orange/ethidium bromide was used to study the effects on apoptosis. *In vitro* wound healing assay and Western blotting were carried out to study effect on cell migration and PI3K–signalling pathway. The results revealed that oxypeucedanin hydrate monoacetate inhibited Caco-2 cell proliferation and decreased the number of cancer colony forming cells. This compound exerted anti-migratory effects in dose- and time-dependent manner. As compared to the untreated control, drug-treated cells showed significant inhibition of cancer cell migration. Oxypeucedanin hydrate monoacetate significantly down-regulated the expression of pAkt and pPI3K in Caco-2 cells.

Introduction

Malignant cancer disorders constitute one of the leading causes of death in most countries. The process of carcinogenesis is a complex biochemical process involving several signalling pathways. So, understanding of these underlying cellular pathways constitutes one of the effective and novel strategies to target uncontrolled cell division in tumor tissues.

Focus has been put on finding novel chemotherapeutic agents which can interfere with these cellular processes and thus inhibit tumor growth and eventually can be used as an anti-cancer drug (Jemal et al., 2011). Colon cancer is ranked as 3rd leading causes of cancer-related mortality in the western world. In the developing

countries, the prevalence of colon cancer is on the rise due to many key factors including changing food habits coupled with increasing life expectancy. As such, colon cancer is a serious public threat throughout the globe and needs urgent attention (Lin et al., 2007). There are various factors which give rise to colon cancer and these include mutations in adenomatous polyposis coli, loss in tumor suppressor gene p53, disruption in cell cycle and suppression in apoptosis (Kinzler and Vogelstein, 1996; Samowitz et al., 2002; Sebolt-Leopold et al., 1999; Torrance et al., 2000). It follows therefore, agents which can manage these above mentioned defects could as potential chemotherapeutic agents in controlling the growth of colon cancer cells.

The objective of the current research work was to study



the antitumor effect of oxypeucedanin hydrate monoacetate isolated from the ethyl acetate extract of *Angelica dahurica* against Caco-2 colon cancer cells by studying its effect on apoptosis, cancer cell migration and PI3K-signalling pathway.

Materials and Methods

Plant material

The roots of the plant were collected from Zhengzhou City, China and identified by Prof. Ying Yang. A voucher specimen (Voucher specimen number: 19-087-011-15) was deposited in the Herbarium of Southeast University, Nanjing, China.

Extraction and isolation of oxypeucedanin hydrate monoacetate

The air dried, finely powdered roots of the fruit (10 kg) was extracted for 48 hours with ethyl acetate in a soxhlet apparatus to yield the extract, which was concentrated under reduced pressure. The ethyl acetate extract (80 g) was loaded on silica gel (60-120 mesh, 500 g) column and eluted with an increasing gradient of petroleum ether and chloroform. Fractions of 100 mL volume each were collected and pooled according to TLC pattern.

Three major fractions were collected (80:20, 60:40, 40:60). The fraction (petroleum ether: chloroform, 60:40) yielded oxypeucedanin hydrate monoacetate as colorless crystalline solid, showing molecular ion peak in the mass spectrum at m/z : 346 analyzed for $C_{18}H_{18}O_7$. The IR spectrum showed prominent peak at 3400 cm^{-1} (-OH), 1738 and 1244 cm^{-1} (-OCOCH₃), 1716 (α , β -unsaturated δ -lactone), 1603 and 1497 cm^{-1} (aromatic), 1382 and 1367 cm^{-1} (gemdimethyl). In its ^1H NMR spectrum, displayed a resonance signal at δ 2.06 (3H, s, C-2'' acetate protons) and down field resonance signals due to olefinic protons at δ 6.20 (1H, d, $J=9.0\text{ Hz}$, H-3), 8.14 (1H, d, $J=9.0\text{ Hz}$, H-4), 7.60 (1H, d, $J=2.2\text{ Hz}$, H-2' furano α -H), 6.91 (1H, d, $J=2.2$, H-3' furano β -H), besides a signal at δ 7.07 (1H, s) due to an aromatic proton (H-8).

Further, the ^1H NMR spectra of the compound displayed the resonance signals at δ 4.59 (2H, m, H-1''), 5.34 (1H, m, H-2''), besides broad signals at δ 3.71 (1H, bs, 3''-OH, exchangeable with D₂O). The signal at δ 1.33 (6H, s) was assigned to gemdimethyl protons. The chemical shift of gemdimethyl indicates that these are attached to carbon, bearing oxygen function. Also, the downfield resonance signal at δ 4.59 displayed by two protons at C-1'' indicates that C-1'' is attached to the aromatic ring through oxygen atom.

In the ^{13}C NMR spectrum, 18 carbon signals were observed. Eleven carbons were assigned for a furanocoumarin nucleus at δC 160.4 (-OC=O, C-2), 112.5

(=CH, C-3), 140.1 (=CH, C-4), 106.2 (=C, C-4a), 149.2 (=C-O-, C-5), 113.6 (=C, C-6), 157.8 (=C, C-7), 94.3 (=C-, C-8), 145.7 (=C-O-, C-8a), 144.6 (=CH, C-2'), and 105.3 (=CH, C-3').

Cell line, culture conditions and MTT cell viability assay

Caco-2 human colon cancer cells were obtained from Shanghai Institute of Cell Resource Center of Life Science (China). Cells were grown in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% fetal bovine serum and 100 U/mL penicillin and 100 $\mu\text{g/mL}$ streptomycin. Cells were cultured in CO₂ incubator with an internal atmosphere of 95% air and 5% CO₂ gas and the cell lines were maintained at 37°C. In order to evaluate the effect of oxypeucedanin hydrate monoacetate on the cell proliferation, MTT assay was performed. Cells were seeded in 96-well plates (1×10^6 cells/well). The cells were treated with the various doses of the compound (0, 4, 12, 24, 48, 96 and 150 μM) for 24, 48 and 72 hours time intervals. Subsequently, MTT solution (5 mg/mL) was then added to each well. After 4 hours incubation time, formazan precipitate was dissolved in 500 μL dimethyl sulfoxide, and then the absorbance was measured in automated microplated reader (Bio-Tek, USA) at 570 nm. The cell viability ratio was calculated by the following formula:

$$\text{Inhibitory ratio (\%)} = (\text{OD control} - \text{OD treated}) / \text{OD control} \times 100\%$$

Cytotoxicity was expressed as the concentration of oxypeucedanin hydrate monoacetate inhibiting cell growth by 50% (IC₅₀ value).

Colony formation assay

Cells were suspended in 1 mL of Dulbecco's modified Eagle's medium containing 0.5% agarose (Amresco, USA) and 10% FBS, and plated on a bottom layer containing 0.8% agarose and 10% fetal bovine serum in 6-well plate in triplicate. After 4 days, plates were stained with 0.3% gentian violet and the colonies were counted under light microscope.

In vitro wound healing assay for studying cancer cell migration

In vitro wound healing assay was performed as per reported method (Liang et al., 2007). Caco-2 colon cancer cells (1×10^6 cells/mL) were seeded in a 6-well plate and incubated at 37°C until 100% full confluent monolayer was achieved. Following 10 hours of starvation, a 250 μL pipette tip was used to produce a straight cell-free wound. Each well was washed twice with PBS to remove any debris and then exposed to numerous concentrations of oxypeucedanin hydrate monoacetate (0, 24, 48 and 150 μM) in a medium. After 48 hours of incubation, the cells were fixed and stained with 3% ethanol comprising 0.5% crystal violet powder for 30

min, and arbitrarily chosen fields were photographed under a light microscope (Olympus, USA). The number of cells that migrated into the scratched area were counted.

Morphological study of apoptosis using fluorescence microscopy and AO/EB staining

Cells were seeded at a concentration of 3×10^5 cell/mL in a volume of 2 mL on sterile cover slip in 6-well tissue culture plates. Subsequent to incubation, the medium was removed and replaced with fresh medium plus 5% FBS and supplemented with oxypeucedanin hydrate monoacetate (0, 24, 48 and 150 μ M). Following the drug treatment, the cover slip with monolayer cells was inverted on the glass slide with 10 μ L of AO/EB stain (50 μ g/mL). The staining images were recorded using a UV fluorescence microscope (Olympus, Japan) using UV filter at 400x magnification to detect morphological evidence of apoptosis.

Western blot assay

Briefly, after cells were treated with oxypeucedanin hydrate monoacetate, the total proteins were extracted with RIPA lysis buffer containing 1% cocktail and 1% phenylmethane sulfonylfluoride (PMSF). Protein concentrations were measured by Bio-Rad protein assay kit (Bio-Rad, USA). Equivalent amounts of proteins were separated by 8–12% sodium dodecyl sulfate polyacrylamide gel electrophoresis and then electrotransferred to a nitrocellulose membrane. After washing with TBST, the membrane was incubated with the corresponding secondary antibodies (1:10000). To probe for pPI3K, Akt, pAkt, Bcl-2, Bax, Bcl-xL, Bad, Cytochrome C, cleaved caspase-3, cleaved-PARP, and β -actin, membranes were incubated overnight at 4°C with the relevant antibodies. Signals were detected

using an ECL Plus Chemiluminescence kit (Applygen Technologies Inc., China) on X-ray film.

Statistical analysis

All experiments were done in triplicate and the results are conveyed as the mean \pm standard deviation. Graphpad Prism (Graphpad Software, Inc., USA) was used for performing statistical analyses and $p < 0.05$ was taken to designate a statistically significant difference.

Results

Anti-tumor activity against Caco-2 colon cancer cells

MTT assay revealed that oxypeucedanin hydrate monoacetate showed a potent growth inhibitory effect on the proliferation of Caco-2 cancer cells with IC_{50} values 36.4, 42.1 and 46.3 μ M at 72, 48 and 24 hours time intervals respectively. Clonogenic assay revealed that oxypeucedanin hydrate monoacetate also leads to a significant reduction in the number of cancer colony forming cells which showed strong dose-dependence (Figure 1 and 2).

Inhibits colon cancer cell migration

Figure 3 shows the effect of oxypeucedanin hydrate monoacetate on the cell migration in Caco-2 cells, indicating that this compound exerts anti-migratory effects in a dose dependent manner. The cells were observed every 12 hours under the same observation field for wound healing position. After 12, 24 and 48 hours time intervals and after 24, 48 and 150 μ M compound doses, the cell migratory abilities decreased considerably as compared to DMSO control. In oxypeucedanin hydrate monoacetate-treated cells, the wound area was larger than that of control (DMSO).

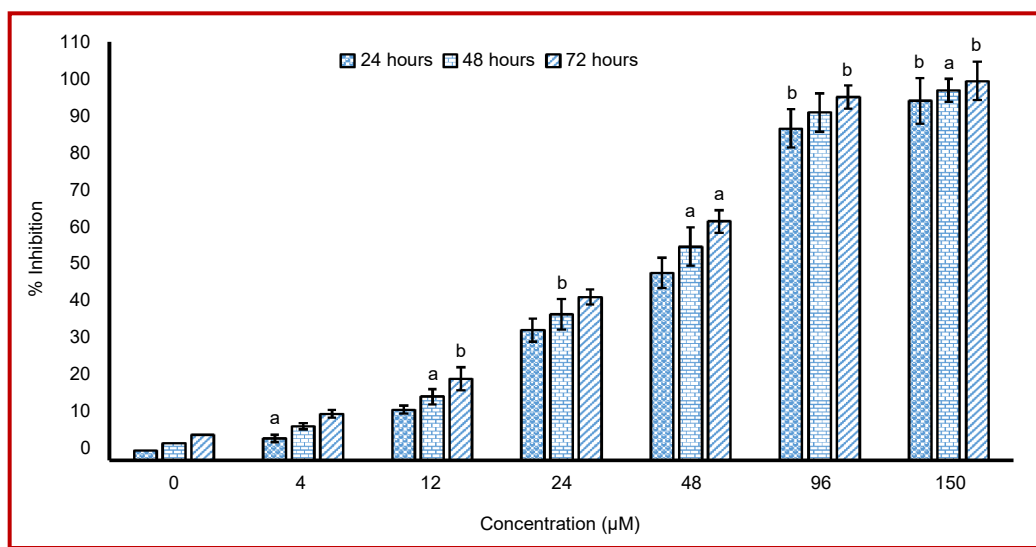


Figure 1: Dose- and time-dependent cytotoxic effect of oxypeucedanin hydrate monoacetate in human colon cancer cells (Caco-2). Data are shown as the mean \pm SD of three independent experiments. ^a $p < 0.05$, ^b $p < 0.01$, vs 0 μ M (control)

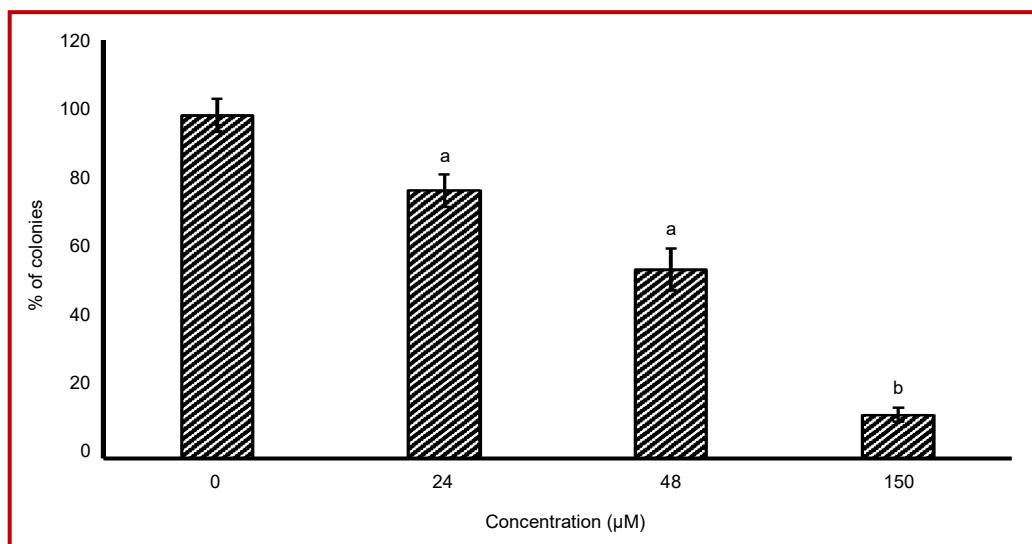


Figure 2: Inhibitory effect of oxypeucedanin hydrate monoacetate on human colon cancer cells (Caco-2). The colony formation assay of Caco-2 cells treated with oxypeucedanin hydrate monoacetate at indicated doses

Induces apoptosis in Caco-2

The results revealed that oxypeucedanin hydrate monoacetate treatment at various doses showed apoptotic changes with yellow green spots in Caco-2 cell nuclei. Higher doses of the compound led to late apoptotic events characterized by orange staining with chromatin condensation and appearance of apoptotic bodies (Figure 4). These morphological changes are the characteristic features of apoptosis process.

Induces apoptosis in Caco-2 cells via PI3K/Akt pathway

It was observed that oxypeucedanin hydrate monoacetate significantly down-regulated the expression of

pAkt and pPI3K in Caco-2 cells in a dose-dependent manner although total Akt protein levels remained constant during all treatments (Figure 5).

Discussion

The results of the current study indicate that oxypeucedanin hydrate monoacetate induced growth inhibitory effects on the proliferation of Caco-2 human colon cancer cells in a dose- and time-dependent manner. Colony formation assay showed that the compound led to a reduction in the number of cancer colony forming cells. *In vitro* wound healing assay revealed that after 12, 24 and 48 hours time intervals and after 24, 48 and

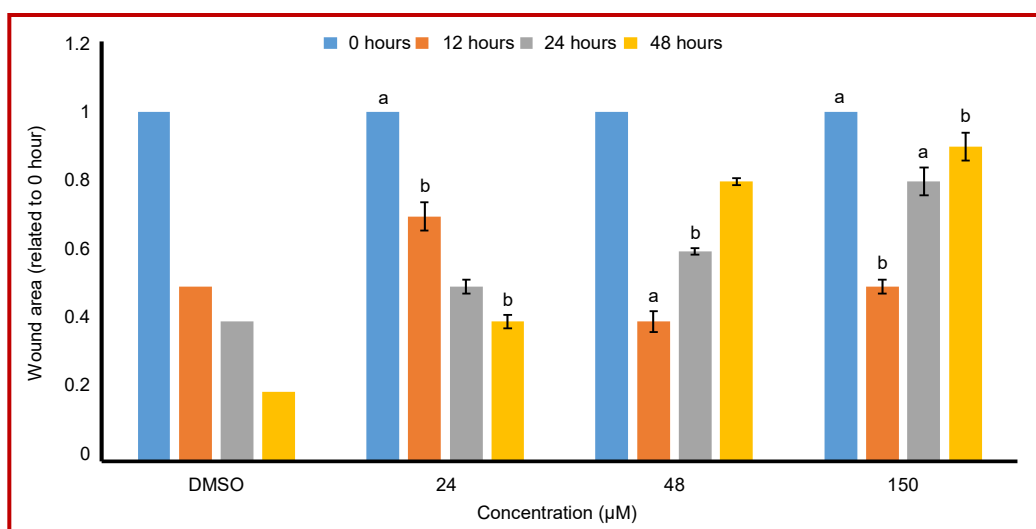


Figure 3: Oxypeucedanin hydrate monoacetate inhibits cell migration in the human colon cancer cells (Caco-2). A wound healing assay was done to determine the cell migration ability. Caco-2 cells were treated with 0.2% DMSO or 24, 48 and 150 µM oxypeucedanin hydrate monoacetate for 0, 12, 24 and 48 hours

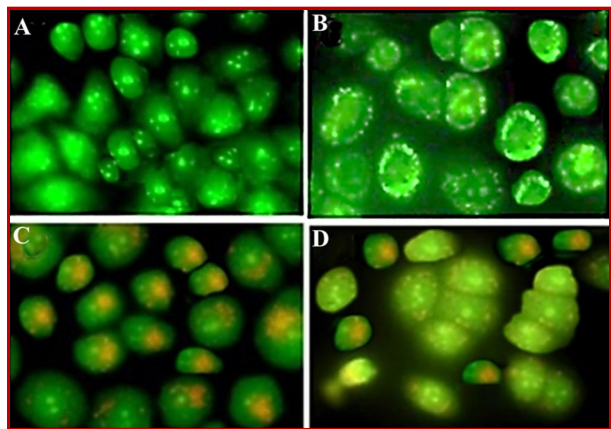


Figure 4: AO/EB staining of Caco-2 human colon cancer cells using fluorescence microscopy. After treating Caco-2 cells with different doses of oxypeucedanin hydrate monoacetate for 48 hours cells were stained and observed under fluorescence microscope. As compared to untreated control group (A), compound-treated cells (B-D) showed clear signs of early and late apoptosis. B, C and D represent effect of 24, 48 and 150 μM dose of oxypeucedanin hydrate monoacetate

150 μM compound doses, the cell migratory abilities decreased considerably as compared to DMSO control. In oxypeucedanin hydrate monoacetate-treated cells, the wound area was larger than that of control (DMSO). This indicated that the compound has the ability to suppress that migratory tendency of cancer cells and hence inhibit cell migration. Further, it was demonstrated that the growth inhibitory effects of oxypeucedanin hydrate monoacetate were mediated via the induction of apoptosis using fluorescence microscopy and AO/EB staining in Caco-2 colon cancer cells. The results showed that lower doses of the compound induced early apoptotic effects characterized by yellow-

ish staining while as higher doses of the compound induced late stage apoptosis marked by orange staining. Finally, the effect of the compound on a crucial signalling pathway known as the PI3K/Akt pathway were observed. This pathway has been known to have a key role in tumorigenesis process and apoptosis. It was observed that oxypeucedanin hydrate monoacetate significantly down-regulated the expression of pAkt and pPI3K in Caco-2 cells in a dose-dependent manner.

Inducing apoptosis in cancer cells is one of the main features of many anti-cancer drugs leading to cancer prevention via controlling cell death. Morphological features of apoptosis include chromatin condensation and nuclear margination (Hale et al., 1996; Kerr et al., 1972; Fadeel et al., 1999; Reed and Tomaselli, 2000). Migration and invasion of malignant cancers are two key features resulting in high morbidity. These are the features of cancer progression and metastasis. It has earlier been reported that cell migration is regulated by several signalling pathways including PI3K, p38MAPK, pJNK and FAK (Lee et al., 2008; Neudauer and McCarthy, 2003). Consequently, research efforts in this direction can result in the therapeutic methods for preventing cancer invasion, migration and metastasis ultimately resulting in improvement of the survival of the cancer patients. Apoptosis is a highly specialized biochemical process which eradicates redundant cells from the body and as such is the key for maintaining tissue homeostasis. Any disturbance in this process eventually results in numerous diseases including cancer. Both intrinsic as well as extrinsic stimuli can trigger the process of apoptosis which finally lead to the activation of proteases (caspases) and nucleases, resulting in destruction of the cell (Adams and Cory, 2007; Cory and Adams, 2002). Oxypeucedanin hydrate mono-

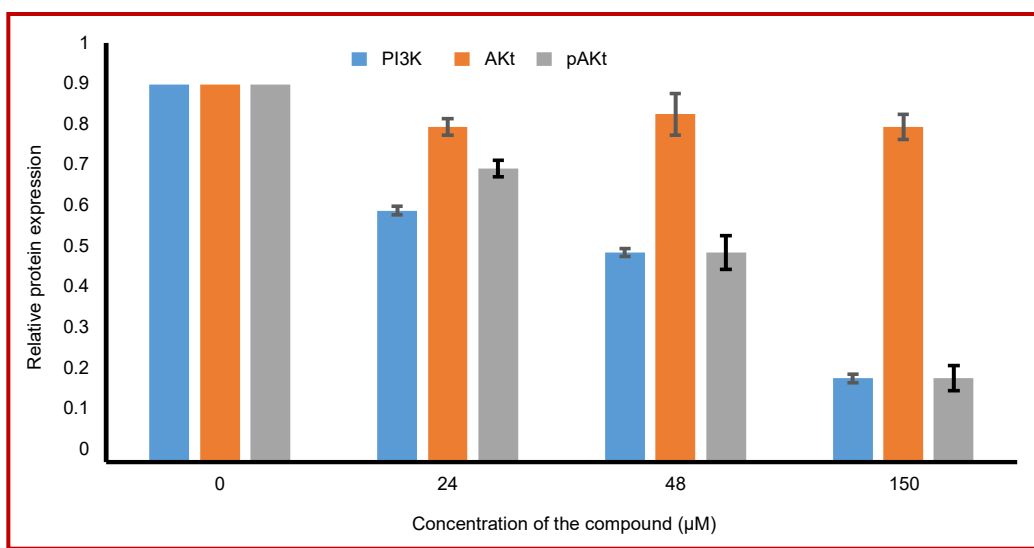


Figure 5: Effects of oxypeucedanin hydrate monoacetate on expression levels of pPI3K, Akt, pAkt. The Caco-2 cells were exposed to 0, 24, 48 and 150 μM dose of the compound for 48 hours and after which whole cell lysates were extracted and subjected to western blotting. The results represent the mean \pm S.D. of at least three independent experiments

acetate induced potent apoptosis in Caco-2 cancer cells as revealed by fluorescence microscopy. Literature survey revealed that oxypeucedanin hydrate has shown cytotoxic activity against epidermoid carcinoma cells (A-431 cells) (Kawai et al., 2001).

Conclusion

The anti-cancer effects of oxypeucedanin hydrate mono-acetate isolated from *A. dahurica* are mediated via the induction of apoptosis, PI3K pathway and suppression of cancer cell migration in Caco-2 human colon carcinoma cells.

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Conflict of Interest

Authors declare no conflict of interest

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