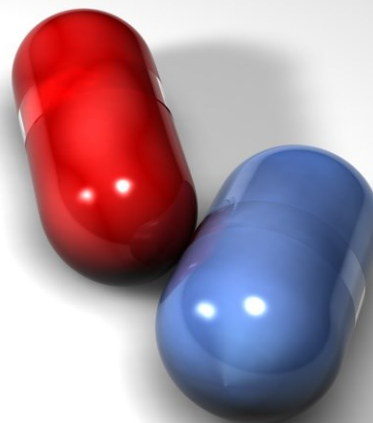


Bangladesh Journal of Pharmacology

Volume: 13; Number 1; Year 2018



Cite this article as: Abdullah S, Ling YS, Daim SJ, Alexander A, Chong KP. *Ganoderma boninense* isolated from Sabah, Malaysia exhibits potent antibacterial activity against clinically important bacterial pathogens. Bangladesh J Pharmacol. 2018; 13: 10-12.



Letter to the Editor

***Ganoderma boninense* isolated from Sabah, Malaysia exhibits potent antibacterial activity against clinically important bacterial pathogens**

Sir,

Ganoderma boninense is a white rot fungus, which mostly can be found in oil palm estates in Southeast Asia. It is an economically devastating pathogen causing major losses on oil palm's profit.

To date, numerous research have been done on *G. boninense*, but they are mainly focused on developing effective control tools in the form of chemicals or biological control agents (Alexander et al., 2017a), study on oil palm defence mechanism (Azura et al., 2016), the fungal pathogenesis (Alexander et al., 2017b), early detection (Alexander et al., 2014), and the molecular

studies (Chong et al., 2011). Meanwhile, there are increasing number on other research field of *G. boninense* which include the study on the fungal metabolites (Alexander et al., 2014) and on pharmaceutical properties (Ismail et al., 2014; Ma et al., 2014). However, despite of all the works done on *G. boninense*, to date, there is no single report on the fungal antibacterial activity against clinically important bacterial pathogens such as *Escherichia coli*, *Klebsiella pneumoniae*, *Pseudomonas aeruginosa*, *Staphylococcus aureus*, *Salmonella enterica*, and *Streptococcus pyogenes* that are increasingly contribute to nosocomial-caused infections.

In the present work, we have isolated a fungal fruiting body (Figure 1A) compromising an oil palm tree in local oil palm plantation in Sabah, Malaysia. The fruiting body was cultured in our laboratory and molecularly identified according to the protocols described by Chong et al. (2011). Molecular analysis revealed that the fungal fruiting body is belong to *G.*

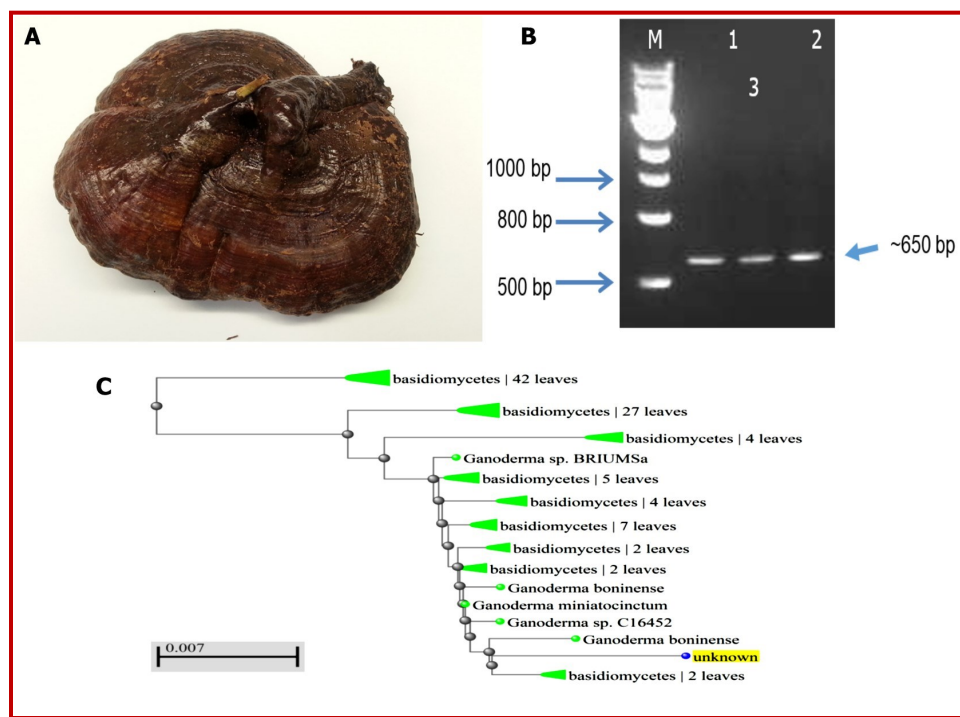


Figure 1: A) The fungal fruiting body obtained from oil palm tree; B) Polymerase Chain Reaction (PCR) analysis of the fungal fruiting body's DNA based on method described by Chong et al. (2011) shows that the PCR product is around 650 bp; C) Phylogenetic Tree analysis (using Basic Local Alignment Search Tool, BLAST, NCBI) of the fungal PCR product DNA sequence (highlighted in yellow) shows that the fungi identified as *G. boninense* species



Table I
Antibacterial activity of *G. boninense* extracts against clinical isolates of bacterial pathogens

Strain†	Zone of inhibition (mm)					
	Aqueous extract#	Methanol extract#	Acetone extract#	Chloroform extract#	Hexane extract#	Tetracycline
1	Not detected	7.7 ± 0.6 ^{cd}	7.7 ± 0.6 ^{cd}	9.3 ± 0.6 ^{fg}	Not detected	17.3 ± 1.5
2	Not detected	Not detected	Not detected	7.0 ± 0.0 ^{ab}	Not detected	15.7 ± 0.6
3	Not detected	Not detected	Not detected	7.0 ± 0.0 ^{ab}	Not detected	15.3 ± 0.6
4	Not detected	8.0 ± 0.0 ^{de}	7.7 ± 0.6 ^{cd}	11.3 ± 0.6 ⁱ	Not detected	23.7 ± 1.5
5	Not detected	7.7 ± 0.6 ^{cd}	Not detected	7.3 ± 0.6 ^{bc}	Not detected	17.3 ± 1.5
6	Not detected	7.7 ± 0.6 ^{cd}	7.7 ± 0.6 ^{cd}	9.7 ± 0.6 ^{gh}	Not detected	21.7 ± 1.5
7*	Not detected	7.0 ± 0.0 ^{ab}	Not detected	7.7 ± 0.6 ^{cd}	Not detected	15.7 ± 1.6 ^k
8*	Not detected	Not detected	Not detected	Not detected	Not detected	11.3 ± 1.6 ^{fgi}
9*	Not detected	Not detected	Not detected	Not detected	Not detected	12.0 ± 1.6 ^{gij}
10*	Not detected	7.3 ± 0.6 ^{bc}	7.7 ± 0.6 ^{cd}	9.3 ± 0.6 ^h	Not detected	16.0 ± 1.6
11*	Not detected	7.3 ± 0.6 ^{bc}	Not detected	7.3 ± 0.6 ^{cd}	Not detected	14.7 ± 1.6 ^k
12*	Not detected	7.3 ± 0.6 ^{bc}	7.0 ± 0.0 ^{ab}	8.7 ± 0.6 ^{fg}	Not detected	13.3 ± 0.6 ^{jk}

†Standard bacterial strains = 1) *E. coli*, ATCC 35218; 2) *K. pneumoniae*, ATCC 1705; 3) *P. aeruginosa*, ATCC 9027; 4) *S. aureus*, ATCC 25923; 5) *S. Enterica*, ATCC 14028, 6) *S. pyogenes* ATCC 19615, and *clinical bacterial isolates = 7) *E. coli*; 8) *K. pneumoniae*; 9) *P. aeruginosa*; 10) *S. aureus*; 11) *S. Enterica*; 12) *S. pyogenes*. #Extracts are in crude form; concentrations and the extracts amount loaded into the disc were standardized to 2 mg/mL and 100 µg respectively. Pure culture of the *G. boninense* fruiting body was obtained according to Chong et al. (2011). The amount loaded into each disc is 30 µg with concentration 1 mg/mL. ND= Antibacterial activity was not detected. None of the clinical bacterial isolates is Tetracycline resistant. The inhibition data are statistically different at $p < 0.05$ unless stated with same letter

boninense species as shown in Figure 1B and Figure 1C. Remarkable antibacterial activity from the identified *G. boninense* extracts was observed against the standard and clinical bacterial isolates as summarized in Table I. Chloroform extract of *G. boninense* gives the broadest spectrum of antibacterial activity against both standard and clinical isolates of bacterial pathogens. For standard bacterial isolates, the greatest inhibition was observed on *S. aureus*, ATCC 25923 (11.3 ± 0.6 mm), followed by the inhibition on *S. pyogenes*, ATCC 19615 (9.7 ± 0.6 mm) and *E. coli*, ATCC 35218 (9.3 ± 0.6). The weakest activity was observed against *K. pneumoniae*, ATCC 1705 and *P. aeruginosa*, ATCC 9027 (7.0 ± 0.0 mm size of inhibition for both isolates). Meanwhile, for the clinical bacterial isolates, the greatest inhibition was observed on *S. aureus* (9.3 ± 0.6 mm size of inhibition) followed by *S. pyogenes* (8.7 ± 0.6 mm size of inhibition) and *E. coli* (7.7 ± 0.6 mm size of inhibition). No antibacterial activity was observed on water and hexane extracts of *G. boninense* against both the standard and clinical bacterial isolates, while two clinical bacterial isolates, *K. pneumoniae* and *P. aeruginosa* were not susceptible to any of *G. boninense* extracts but to tetracycline. From this work, we found that the chloroform and methanol extracts of *G. boninense* by some part give broader spectrum of antibacterial activity against the tested bacterial pathogens compared to other extracts. This work also suggest that *G. boninense* might bearing potent antibacterial agent against important nosocomial infections-related bacterial pathogens, but right solvents system

and extraction procedures are crucial to extract them out. One of *Ganoderma* species, *G. lucidum* is well known to exhibit potent medicinal potential including antibacterial activity (Iftekhara et al., 2011). This report confirmed that *G. boninense*, as wood decaying fungi like *G. lucidum* can also exhibit potent antibacterial potential. In-depth investigation is necessary to identify the responsible antibacterial compounds and the exact antibacterial mode of action against the selected bacterial pathogens.

The authors acknowledge their profound gratitude to Ministry of Education Malaysia for financially supporting the research through Fundamental Research Grant Scheme (FRG0348), Faculty of Science and Natural Resources, Faculty of Medicine and Health Sciences, and Biotechnology Research Institute of Universiti Malaysia Sabah for providing the facilities for research work.

Syahriel Abdullah¹, Yee Soon Ling², Sylvia Jerome Daim³, Arnyitte Alexander¹, Khim Phin Chong¹

¹Biotechnology Programme, Faculty of Science and Natural resources, Universiti Malaysia Sabah, Jalan UMS, 88400, Kota Kinabalu, Sabah, Malaysia; ²Biotechnology Research Institute, Universiti Malaysia Sabah, Jalan UMS, 88400, Kota Kinabalu, Sabah, Malaysia; ³Microbiology and Pathobiology Department, Faculty of Medicine and Health Sciences, Universiti Malaysia Sabah, Jalan UMS, 88400, Kota Kinabalu, Sabah, Malaysia.

Corresponding author:
 email: chongkp@ums.edu.my; Tel.: +6088-320000 Ext 5655;
 Fax: +6088-435324

- Alexander A, Abdullah S, Rossall S, Chong KP. Evaluation of the efficacy and mode of action of biological control for suppression of *Ganoderma boninense* in oil palm. *Pakistan J Bot.* 2017a; 49: 1193-99.
- Alexander A, Phin CK. Combination of biological agents in suppressing colonization of *Ganoderma boninense* of basal stem rot. *Am-Eurasian J Sustain Agric.* 2014; 8(Spec Issue 2): 1-7.
- Alexander A, Sipaut CS, Dayou J, Chong KP. Oil palm roots colonisation by *Ganoderma boninense*: An insight study using scanning electron microscopy. *J Oil Palm Res.* 2017b; 29: 262-66.
- Azura AN, Yusoff M, Tan GY, Jegadeesh R, Appleton DR, Vikineswary S. *Streptomyces sanglieri* which colonised and enhanced the growth of *Elaeis guineensis* Jacq. seedlings was antagonistic to *Ganoderma boninense* in *in vitro* studies. *J Ind Microbiol Biotechnol.* 2016; 43: 485-93.
- Chong KP, Lum MS, Foong CP, Wong CM, Atong M, Rossall S. First identification of *Ganoderma boninense* isolated from Sabah based on PCR and sequence homology. *Afr J Biotechnol.* 2011; 10: 14718-23.
- Iftekhhar AM, Choudhry ZK, Khan MI, Saleh AA. Comparative study of antibacterial activity of wood-decay fungi and antibiotics. *Bangladesh J Pharmacol.* 2011; 6: 14-17.
- Ismail K, Abdullah S, Chong KP. Screening for potential antimicrobial compounds from *Ganoderma boninense* against selected food borne and skin disease pathogens. *Int J Pharm Pharm Sci.* 2014; 6: 771-74.
- Ma K, Ren J, Han J, Bao L, Li L, Yao Y, Sun C, Zhou B, Liu H. Ganoboninketals A-C, antiplasmodial 3, 4-seco-27-norlanostane triterpenes from *Ganoderma boninense* Pat. *J Nat Prod.* 2014; 77: 1847-52.
-