

## VISUAL EXPERIMENT

## Estimation of vitamin B<sub>12</sub> in plasma by High Performance Liquid Chromatography

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### ABSTRACT

High performance liquid chromatography was used to develop and validate the detection of vitamin B<sub>12</sub> in blood plasma sample. The mobile phase consists of a mixture of 0.025% trifluoroacetic acid in deionized water and 30% methanol. The mixture used was adjusted to pH 2.9 and the flow rate was adjusted to 0.5 mL/min. The separation was achieved using C18 column maintained at 30°C temperature and detection of vitamin B<sub>12</sub> was conducted at maximum wavelength 230 nm.

### INTRODUCTION

Vitamin B<sub>12</sub> (cobalamin and its derivatives) is an essential water soluble micronutrient that plays an important role in regulation of brain functioning (Grotzkj et al., 2012). The deficiency of vitamin B<sub>12</sub> may lead to various complications such as neurological abnormalities like peripheral neuropathy, autonomic dysfunction, psychosis, memory impairment, Cognitive decline etc (Roynolds et al., 2006). The normal range of vitamin B<sub>12</sub> in plasma of healthy subject is 200-900 pg/mL.

Vitamin B<sub>12</sub> in plasma, serum and urine can be measured by radioimmunoassay, enzyme-linked immunosorbent assay, microbiological assay (using *Lactobacillus leichmannii*; Girdwood, 1954), HPLC (Stefova et al., 1997), capillary electrophoresis, mass spectroscopy, Raman spectroscopy (Mayer et al., 1973), Fourier transform Raman spectroscopy (Hancewicz and Petty, 1995), chemiluminescence (Song and Hou, 2003) and fluorescence quenching.

A deficiency vitamin B<sub>12</sub> occurs when the level is less than 200 pg/mL (Shaik and Gan, 2013). Because of its very low concentration in plasma, it is very challenging to develop a method with good sensitivities and specificities. Currently, the available methods to estimate vitamin B<sub>12</sub> in plasma are mainly based on microbiological assay which is time consuming, hazardous to the health of the operator and may lack specificity (Giorgi et al., 2012). Different analytical methods have been developed over recent years due to increasing demand of vitamins. However, most of these methods were applied in food particle, drinks, and vitamin supplements and so on.

There are very few analytical methods developed to detect vitamin B<sub>12</sub> in plasma due to extensive time consumption in sample preparation, robustness and reproducibility.

Therefore, the aim of this study was to develop and validate a novel High Performance Liquid Chromatography (HPLC) methodology for rapid detection and quantitation of water soluble vitamin B<sub>12</sub> in biological fluids (plasma) which can lead to both time and cost saving.

### MATERIALS AND EQUIPMENTS

1. HPLC unit
2. Analytical balance
3. Sonicator (Sonorex Super 10 P)



4. Centrifuge (Labofuge 200)
5. Nitrogen/ Argon gas cylinder
6. Filter paper (0.2  $\mu\text{m}$ ) for filtration
7. Microtip pipette (Thermo Scientific)
8. Vortex mixer (VM 2000 DIGI System)
9. pH paper (Fisher brand FB 33041)
10. Vitamin B<sub>12</sub> (Sigma-Aldrich)
11. Dichloromethane (Merck. KGa, Germany)
12. Trifluoroacetic acid (Scharlab S.L, Spain)
13. Methanol (HPLC grade)
14. Deionized water

### PREPARATION OF REAGENTS

**Mobile phase:** Solution A is prepared by adding trifluoroacetic acid (100  $\mu\text{L}$ ) into 400 mL of deionized water. The pH is adjusted to 2.9. Solution B consists of 150 mL methanol. Then 350 mL of solution A and 350 mL of solution B are mixed to make a final volume of 500 mL. Further, 500 mL of this solution is sonicated for 2 min before using as mobile phase. A fresh solution is prepared daily.

**Standard solution of vitamin B<sub>12</sub>:** Weigh 1 mg of vitamin B<sub>12</sub> into a test tube and dissolve it in 1 mL of water with TFA (1 mg/mL). Then make serial dilution into 100  $\mu\text{g/mL}$ , 10  $\mu\text{g/mL}$ , 1  $\mu\text{g/mL}$  and 100 ng/mL.

### VIDEO CLIP

Vitamin B<sub>12</sub> assay: 4 min 9 sec

### PROTOCOL

#### Preparation of the Sample

1. Take 400  $\mu\text{L}$  of fresh plasma in a test tube
2. Add 800  $\mu\text{L}$  of dichloromethane to precipitate protein
3. Vortex the sample for 30 sec
4. Centrifuge the sample at 3,500 rpm for 10 min to separate the precipitate from the supernatant
5. Collect the supernatant into another test tube
6. Add 400  $\mu\text{L}$  of methanol to the supernatant
7. Wait for 10 min
8. Vortex the sample for 30 sec
9. Centrifuge the sample at 3,500 rpm for 10 min again to observe the two clear layer of solution
10. Separate the upper layer into another test tube
11. Evaporate the methanol using nitrogen/argon gas
12. Add 200  $\mu\text{L}$  of solution A into the test tube to dissolve vitamin B<sub>12</sub>
13. Vortex the sample for 30 sec

14. Adjust the pH of the sample to 2.9
15. Pour the sample/standard solution into a HPLC vial

### Set-up of HPLC

1. Make sure that bottle contains >400 mL of mobile phase
2. Turn on instruments in the following order: computer, HPLC software, degasser, pump, column compartment, autosampler and detector
3. Set the flow rate to 0.5 mL/min
4. Set the injection volume to 50  $\mu$ L
5. Set the column temperature to 30°C
6. Set the wavelength of UV detector to 230 nm

### Injection of Sample and Data Collection

1. Keep the HPLC vial into the autosampler tray
2. Set the run time using the computer
3. Click the "Start" button
4. Inject 50  $\mu$ L of the sample/standard into the C<sub>18</sub> column [150 (length) x 4.6 mm (internal diameter), particle size 5  $\mu$ m] using an autosampler
5. Data collection can be stopped at any time by either clicking on the Stop button
6. When data collection is complete, either through a manual stop run or when the preset run time is complete, a report will be generated
7. Identify the unknown peak from the standard known peak using the retention time
8. Printer the data in a A4 size paper
9. Turn off the instruments in this order: HPLC software, detector, autosampler, column compartment, pump, degasser and computer

### CALCULATION

1. Identify the retention time of the standard peak
2. Confirm the unknown peak from the sample using the retention time of the standard peak
3. Inject series of vitamin B<sub>12</sub> standard solutions (minimum 4) using the same procedure
4. Record peak areas
5. These four levels should be within the calibration range
6. Calculate the concentration of vitamin B<sub>12</sub> using the following formula:

$$\frac{\text{Peak area of sample}}{\text{Peak area of standard}} \times \text{Concentration of mixed standard solution, pg/mL} \times \text{Dilution volume of sample, mL} / \text{Sample weight, g}$$

### DISCUSSION

In this study, vitamin B<sub>12</sub> was measured by HPLC. In comparison to other sophisticated methods, HPLC method requires sample preparation whereas electroluminescence or radioimmunoassay does not (Karmi et al., 2011). In addition, HPLC method (68,000 pg/mL) is less sensitive than the radioimmunoassay (200 pg/mL) or electroluminescence (30 pg/mL). Cost and availability of the instrument are considered for estimation.

In HPLC, optimization of the method is required. It includes: a) suitable mobile phase, b) wavelength, c) with/without buffer and its type and concentration, d) pH of the mobile phase, e) column temperature, f) injection volume, and g) flow rate.

Vitamin B<sub>12</sub> is a water soluble vitamin. Therefore, mobile phase should be a polar one. Methanol or acetonitrile is preferred.

The wavelength is also important. The reported wavelengths used for the estimation of vitamin B<sub>12</sub> were 210 (Shaik and Gan, 2013), 230 (Giorgi et al., 2012), 260, 254, 360 (Shaik and Gan, 2013), 545 and 550 nm. We used 230 nm. In this study, methanol was used. The UV absorbance cutoff of acetonitrile or methanol is 200 or 205. Therefore, wavelength of 210 should better be avoided.

The pH is important factor for the determination of vitamin B<sub>12</sub>. The peak is marked at low pH. A study was conducted on using mobile phases of different pH such as 2.3 (Shaik and Gan, 2013), 2.6 (Heudi et al., 2005), 2.9 (Giorgi et al., 2012), 4.0 and 6.0 and found that pH 2.3 was the best for vitamin B<sub>12</sub> peak which is good in peak shape and height (Shaik and Gan, 2013). Because sometimes, the peak may be broaden.

At a flow rate of 0.1 mL/min, vitamin B<sub>12</sub> was not eluted (Shaik and Gan, 2013).

## REFERENCES

- Gentili A, Caretti F, D'Ascenzo G, Marchese S, Perret D, Di Corcia D, Rocca LM. Simultaneous determination of water-soluble vitamins in selected food matrices by liquid chromatography/electrospray ionization tandem mass spectrometry. *Rapid Commun Mass Spectrom*. 2008; 22: 2029-43.
- Girdwood RH. Rapid estimation of the serum vitamin B<sub>12</sub> level by a microbiological method. *BMJ*. 1954; 2(4894): 954-56.
- Giorgi MG, Howland K, Martin C, Bonner B. A novel HPLC method for the concurrent analysis and quantitation of seven water-soluble vitamins in biological fluids (plasma and urine): A validation study and application. *Sci World J*. 2012.
- Hancewicz TM, Petty C. Quantitative analysis of vitamin a using Fourier transform Raman spectroscopy. *Spectrochim Acta A Mol Spectrosc*. 1995; 51: 2193-98.
- Heudi O, Kilinc T, Fontannaz P. Separation of water-soluble vitamins by reversed-phase high performance liquid chromatography with ultra-violet detection: Application to polyvitaminated pre-mixes. *J Chromatogr A*. 2005; 1070: 49-56.
- Karmi O, Zayed A, Baraghehi S, Qadi M, Ghanem R. Measurement of vitamin B<sub>12</sub> concentration: A review on available methods. *IIOAB J*. 2011; 2: 23-32.
- Klejduš B, Petrová J, Potěšil D, Adam V, Mikelová R, Vacek J, Kizek R, Kubáň V., Simultaneous determination of water-and fat-soluble vitamins in pharmaceutical preparations by high-performance liquid chromatography coupled with diode array detection. *Anal Chim Acta*. 2004; 520: 57-67.
- Liu BS, Gao J, Yang GL. Determination of vitamin B<sub>12</sub> concentration by fluorescence quenching with acridine orange-rhodamine 6G energy transfer system. *Guang Pu Xue Yu Guang Pu Fen Xi*. 2005; 25: 1080-82.
- Mayer E, Gardiner DJ, Hester RE. Resonance Raman spectra of vitamin B<sub>12</sub> and dicyanocobalamin. *Biochim Biophys Acta Gen Sub*. 1973; 297: 568-70.
- Sami R, Li Y, Qi B, Wang S, Zhang Q, Han F, Ma Y, Jing J, Jiang L. HPLC analysis of water-soluble vitamins (B<sub>2</sub>, B<sub>3</sub>, B<sub>6</sub>, B<sub>12</sub>, and C) and fat-soluble vitamins (E, K, D, A, and  $\beta$ -carotene) of okra (*Abelmoschus esculentus*). *J Chem*. 2014.
- Shabangi M, Sutton JA. Separation of thiamin and its phosphate esters by capillary zone electrophoresis and its application to the analysis of water-soluble vitamins. *J Pharm Biomed Anal*. 2005; 38: 66-71.
- Shaik MM, Gan SH. Rapid resolution liquid chromatography method development and validation for simultaneous determination of homocysteine, vitamins B<sub>6</sub>, B<sub>9</sub>, and B<sub>12</sub> in human serum. *Indian J Pharmacol*. 2013; 45: 159-67.

- Song Z, Hou S. Sub-picogram determination of vitamin B<sub>12</sub> in pharmaceuticals and human serum using flow injection with chemiluminescence detection. *Anal Chim Acta*. 2003; 488: 71-79.
- Stefova M, Stafilov T, Stojanosk, K, Cepreganova-Krstic B. Determination of vitamin B<sub>12</sub> in multivitamin tablets by high performance liquid chromatography. *Anal Lett*. 1997; 30: 2723-31.
- Yin C, Cao Y, Ding S, Wang Y. 2008. Rapid determination of water-and fat-soluble vitamins with microemulsion electrokinetic chromatography. *J Chromatogr A*. 2008; 1193: 172-77.

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**PRECAUTION**

Prepare fresh buffer solution to avoid any bacterial contamination